Synthesis of a Tritiated 3-Dehydroecdysteroid Putative Precursor of Ecdysteroid Biosynthesis in Locusta migratoria

Frédéric DOLLE¹, Charles HETRU², Jean-Pierre ROUSSEL², Bernard ROUSSEAU³ Franck SOBRIO³, Bang LUU^{1*}, Jules A HOFFMANN²

¹Laboratoire de Chimie Organique des Substances Naturelles, associé au CNRS

5 rue Blaise Pascal, 67084 Strasbourg, France

²Laboratoire de Biologie Générale, associé au CNRS, 12 rue de l'Université, 67000 Strasbourg, France

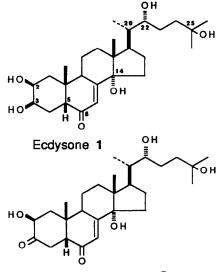
³Service des Molécules Marquées, Departement de Biologie Cellulaire et moléculaire

CE de Saclay, 91191 Gif-sur-Yvette, France

(Received in Belgium 21 May 1991)

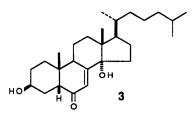
Abstract - We have synthesized a tritiated form of 14α -hydroxy-5 β -cholest-7-ene-3,6-dione (5 β -diketol) of high specific activity (1 74 TBd/mmol) from 7-dehydrocholesterol in seven steps with stereoselective introduction of the A/B cis ring junction as the key reaction. We have examined the ability of endocrine glands (prothoracic glands) of *Locusta migratoria* to use this molecule as a precursor of 3-dehydroecdysone and of ecdysone biosynthesis. A very efficient conversion of 5 β -diketol to ecdysone and to 3-dehydroecdysone was monitored, which opens up new possibilities for the understanding of the role of 3-dehydro-compounds in ecdysteroid biosynthesis

INTRODUCTION



3-Dehydroecdysone 2

Ecdysteroids represent a family of polyhydroxylated steroids which serve as hormonal messengers in insects The parent molecule, ecdysone (1), which is mostly referred to as the molting and metamorphosis hormone, was considered as the hormone synthesized and secreted during postembryonic development by endocrine glands (known as prothoracic glands or homologous structures) Recently it has been reported that, in some insect species, prothoracic glands secrete 3-dehydroecdysone (2) in vitro in addition to ecdysone (1)^{1,2} This molecule is rapidly reduced to ecdysone by a hemolymph reductase which exclusively reduces the keto group to 3β -hydroxyl³ In ecdysone biosynthesis investigations, 3-oxo compounds have never been identified, but the precursors used for these studies contained a C-3 β hydroxy group⁴



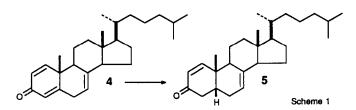
These data prompted us to undertake the synthesis of 14α -hydroxy-5 β -cholest-7-ene-3,6-dione (17, 5 β -diketol) in labelled form This molecule is related to 5 β -ketodiol 3 (3 β ,14 α -dihydroxy-5 β -cholest-7-en-6-one), which is a well established precursor in ecdysone biosynthesis⁵

The instability of the A/B cis ring junction (5 β -H configuration) due to the absence of a 2 β -hydroxy group in the target molecule and the opportunity for the stereoselective hydrogenation of the Δ^4 double bond which will give the desired 5 β configuration, led us to develop a novel route for the synthesis of 2-deoxyecdysteroids

The strategy which we propose in the present paper for the synthesis of labelled 5 β -diketol 17 is firstly the hydrogenation of a Δ^4 double bond to give the A/B cis ring junction and secondly the labelling of the molecule at the last step by tritiation of a Δ^1 double bond.

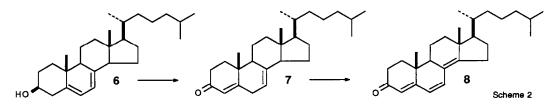
SYNTHESIS OF 14a-HYDROXY-5β-CHOLEST-7-ENE-3,6-DIONE

The key compound of our synthesis is 5, which has all the structural features required to lead to the target molecule 17.1) the A/B cis ring junction is indispensable for the biological activity, ii) the Δ^7 double bond has two allylic sites C-6 and C-14 which can be efficiently oxidized to the conjugated hydroxy enone system; iii) the Δ^1 double bond can be specifically reduced with tritum at the last step



It was likely that compound 5 could be obtained readily from 4 by selective reduction of Δ^4 double bond without any reaction on Δ^1 or Δ^7 double bonds, as described by Rubin and Armbrecht⁶ (Scheme 1)

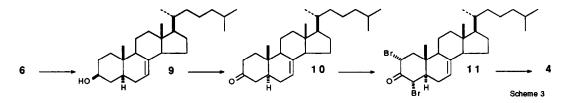
Thus we planned to synthesize compound 4 directly from 7-dehydrocholesterol (6) in a few steps (Scheme 2) The dienone 7 is obtained by Oppenauer oxidation⁷ in good yield (85%)



Treatment of 7 with DDQ $(2,3-dichloro-5,6-dicyano-benzoquinone)^{8,9}$ in refluxing dioxan, unfortunately produced the polyenone 8 in 55% yield instead of the expected product 4

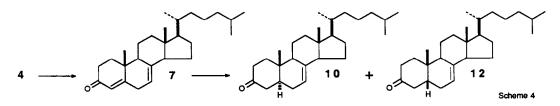
This compound 8 was also obtained during the attempted bromination of 7 with tri-N-methylanilinium perbromide¹⁰ due to spontaneous dehydrobromination

We overcame this difficulty by using a longer synthetic pathway (Scheme 3) 7-Dehydrocholesterol (6) was easily reduced by catalytic hydrogenation with palladium on charcoal. Lathosterol (9), which was obtained with a very good yield (overall 98%), was oxidized with PCC (pyridinium-chlorochromate) adsorbed on alumina¹¹ This technique is very convenient because PCC can be easily eliminated by filtration and the yield was particularly high (95% for compound 10) The ketone 10 was brominated with two equivalents of tri-N-methylanilinium perbromide in THF at 0°C Recristallization gave the pure dibromide in 64% yield The dibromide 11 was fully characterized and the ¹H NMR spectrum indicated that the bromine atoms occupied the α positions (¹H NMR $\delta = 4.67$ ppm, d, J = 13.2 Hz, 1H, H-4_{ax} and $\delta = 4.87$ ppm, dd, J = 5.8 and 14.2 Hz, 1H, H-2_{ax}) Dehydrobromination¹² in refluxing DMF with LiBr and LiHCO₃ afforded the trienone 4 (90%)



Compound 4 was submitted to the well described reduction⁶ with palladium on calcium carbonate in the presence of quinoline and ethanol and afforded a mixture of 7, 10 and 12

It is clear that the Δ^1 double bond was first reduced, followed by reduction of the Δ^4 double bond which gave the ketones 10 and 12 (Scheme 4). The Δ^7 double bond never reacts under these conditions Similarly, reduction with palladium on charcoal (with or without quinoline), with other catalysts and in various solvents, led first to the hydrogenation of the Δ^1 double bond. A recent publication¹³ confirmed this disappointing result.



In consequence we have devised an original regio- and stereoselective synthetic pathway to obtain compound 5 (Scheme 5)

Compound 7 was directly reduced to give the A/B cis ring junction with palladium on charcoal in a mixture of aqueous NaOH 0 1N and ethanol $(85\%)^{14\cdot16}$ The reduction was regioselective, only the Δ^4 double bond being hydrogenated, and stereoselective, the junction of the A/B ring being in cis configuration (H in 5 β) with 82% diastereoisometric excess The A/B cis ring junction was confirmed by ¹H and ¹³C NMR (¹H NMR $\delta = 0.99$ ppm (CH₃-19, cis), $\delta = 1.01$ ppm (CH₃-19, trans), ¹³C NMR $\delta = 23.5$ ppm, 33.9 ppm and 42.0 ppm (C-19, C-9 and C-5 cis), $\delta = 12.5$ ppm, 49.0 ppm and 43.0 ppm (C-19, C-9 and C-5 trans), For references, see Table 1)

The bromination with tri-N-methylanilinium perbromide yielded the 2 β bromoderivative 13 (80%) Attempted recristallization of an aliquot from methanol/ether gave the bromo-ketal 14 The ¹H NMR spectrum indicated that the bromine atom occupied the 2 β position (¹H NMR . δ = 4 34 ppm, dd, J = 4 5 Hz and 13 0

						_											_													
18	152 6*	129 2*	188 1+	126 6*	157 8	185 6+	123 8*	166 3	42 1	43 6	216	306	46 6	853	32 2	266	506	159	169	35 6	216	363	240	39.5	280	22 5-	22 8-			
17	31 2*	34 9*	208 3	37 0*	553	199 4	120 5	164 6	35.5	366	215	39 1	476	85 5	317	266	507	159	22 8	35 6	189	363	24 1	39 5	280	22.5-	22 8-			
16	159 9*	128 8*	198 7+	363	52 0	196 3+	1212	164.5	507	40 9	313	39 1	48 1	85 6	315	26 6	50.7	16.0	22 6	35 6	189	35.6	241	39.5	28 0	22 6-	22 8-			
15	157 7*	128 8*	198 0+	357	52 0	1964+	121 5	164 6	565	39.5	29.7	393	465	566	22 1	278	566	12.4	227	360	188	360	239	39.5	280	22 6-	22 8-			
14	36 9*	39.5*	99.2	45 5*	550	33.8	1151	137.5	37.8	37 1	22 0	39 6	437	53 8	22.8	275	560	119	24 1	36 2	188	36.0	23 9	39.5	280	22 6-	22 8-	1	49.5"	50 8"
12	35 6*	37 8*	212 3	43 6*	42.0	28 5	1146	138 3	38 1	33 9	22 4	40 0	44 1	549	22 8	28 0	564	119	23 5	36 2	189	36.2	240	39 6	28 6	22 6-	22 8-			
11	509	52 2*	193 0	51 5*	59 5*	29 6	1167	138 6	48 8	388	214	39 3	431	546	22 8	278	557	119	141	361	18.7	35 9	237	39.7	280	22 5-	22 8-			
10	38 2*	39 6*	2118	44 3*	43 0	30 2	1170	139 6	49 0	345	218	38 9	43 5	55 1	23 0	280	563	119	125	36.2	189	36.2	240	39 6	28 0	22 6-	22 8-			
6	31 6*	37.3*	713	39 6*	404	298	117.5	139 7	49 6	343	217	38 1	43.5	55 2	23 0	280	564	119	13 1	36.2	189	36.2	240	39 6	28 0	22 6-	22 8-			
80	39.4*	34 1*	199 5	122 9	164 5+	124 4'	134 1'	156 3+	44 3	368	18 8	35.7	4 2	1243	254	272	557	167	189	346	19 0	359	23.7	39.5	280	22 6-	22 8-			-
7	33 4*	34 2*	199 0	122 8	168 6	32 9*	1155	139 6	46 1	38 1	22 1	393	43 6	55 0	22 9	279	562	6 11	212	36.2	189	36.2	240	39 6	28 0	22.5-	22.7-			
5	162 2*	127 8*	2005	40.5	40 2	278	1151	139 3	43 6	376	22.7	39 6	43.9	54.8	22 9	279	563	119	22.2	36 2	189	36 2	240	39 6	28 0	22 6-	22 8-			
4	155 0*	127 9*	1861	123 5*	1659	32.7	1158	138 4	479	424	22 8	39 1	43.5	547	23 0	278	560	119	19.5	361	188	36 1	239	39.5	280	22.5	22.5			
	1	7	3	4	Ś	6	2	~	9	01	П	12	3	14	15	16	17	18	19	ຊ	21	ង	ន	\$	ห	R	27		-	5

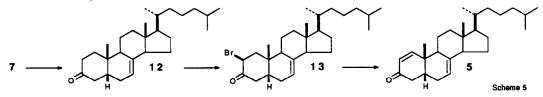
Table 1 : ¹³C NMR chemical shifts

The assignments were based upon (1) shuelding data, (2) by comparison with the spectra of closely related ecdysteroids^{a-c} and steroids^d References (a) W B Smuth. Org. Magn Reson, 1977, 9, 644, (b) T Haag et al. J Labelled Compd Radiopharm, 1985, 22, 547 (c) See ref 18 and ref 21, (d) J W Blunt et al. Org. Magn Reson, 1977, 9, 439

δ_c (50 MHz, standard Me₄St) * (or ⁺, ⁺, ⁺, ⁺) unterchangeable assignment Solvent CDCl₃ (77 02)

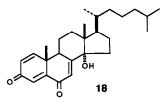
F DOLLE et al

Hz, 1H, H- 2_{ax}) The dehydrobromination was performed in refluxing DMF with LiBr and LiHCO₃ and gave the enone 5 (80% yield)

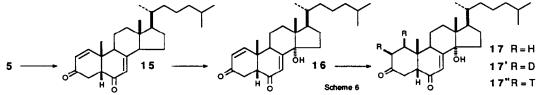


Oxidation of the allylic position C-6 in compound 5 was carried out in dry methylene chloride using freshly prepared¹⁷ CrO₃/(pyridine)₂ complex (Collins reagent), and the dienone 15 was obtained with a moderate yield of 30% (Scheme 6) Oxidation in C-14 α must be performed under strong conditions, because

the A/B cis ring junction introduces a substantial steric hindrance of the α face of the steroid Treatment of the dienone 15 in refluxing dioxan with 10 equivalents of selenium dioxide¹⁸ afforded in a few minutes compound 16 with a 75% yield. By carefully monitoring the reaction by TLC, the formation of 14 α -hydroxy-cholesta-1,4,7-triene-3,6-dione (18, by-product) could be minimized^{9,19}



Selective reduction of the Δ^1 double bond was performed with palladium on charcoal and yielded the expected compound 17 quantitatively. The hindered Δ^7 double bond has never been reduced in our experimental conditions



Together with the mass spectrometric analysis, ¹H NMR spectrometry clearly demonstrated the reduction of the Δ^1 double bond (disappearance of the signals at $\delta = 5.95$ ppm, d, J = 10.0 Hz, 1H, H-2 and $\delta = 6.83$ ppm, d, J = 10.0 Hz, 1H, H-1)

In order to analyse the positions and the number of the hydrogen atoms introduced, we carried out a deuteriation under the conditions which we anticipated to be used in trittation

²H NMR analysis indicated the introduction of two deuterium atoms (coupling constants of ²H at C-1 and at C-2, $\delta = 1.52$ ppm, m, $w_{1/2} = 12.3$ Hz, ²H-1 and $\delta = 2.33$ ppm, m, $w_{1/2} = 14.2$ Hz, ²H-2). The analysis by mass spectrometry of the reaction products gave the following data $D_0.5\%$, $D_1.48\%$, $D_2.47\%$, $D_3.0\%$

To determine the precise configuration of the deuteriums introduced in positions C-1 and C-2, the deuteriated compound 17' has been reduced using sodium borohydride in methanol to give exclusively the 3- α alcohol²⁰ (19) Under these conditions, the coupling constants of the proton at position C-3 β (td, J_{H3ax-H2eq} and J_{H3ax-H4ax} = 11 8 Hz, J_{H3ax-H4eq} = 3 54 Hz, J_{Deq-H3ax} = very weak) were consistent with the incorporation of one deuterium at position C-2 β . As the palladium on charcoal is well known to perform cis additions, the second deuterium should be in position C-1 β

Tritiation was performed at the Commissariat à l'Energie Atomique (Saclay, France) The specific activity of 14α -hydroxy-5 β -cholest-7-ene-3,6-dione (17") was 1 74 TBq/mmole (47 Ci/mmol).

CONVERSION STUDIES OF TRITIATED 14α-HYDROXY-5β-CHOLEST-7-ENE-3,6-DIONE (17")

Prothoracic glands were excised from five larvae of *Locusta* of the mid fifth instar (period of intense ecdysone (1) synthesis *in vivo*) and incubated in modified Landureau's medium²¹ in the presence of $0.2 \,\mu$ M of the newly-synthesized radiolabelled 5 β -diketol 17" After 16h the medium was injected into a reversed phase HPLC and eluted by a methanol/water gradient (fig 1) In this system, ecdysone and 3-dehydroecdysone are known to co-elute²² The radioactivity which clearly co-eluted with reference ecdysone was rechromatographed in a system with higher resolution (fig 2) to separate the presumed ecdysone and 3-dehydroecdysone. The two molecules are in similar proportions. The identity of putative ecdysone was ascertained by co-acetylation with unlabelled ecdysone (data not shown). The identity of putative 3-dehydroecdysone (fractions 30 min - 32 min) was ascertained by a biological approach, as there is not an easy chemical reaction to relate labeled 3-dehydroecdysone to ecdysone. But incubation of the putative 3-dehydroecdysone and 3-dehydroecdysone as conversion products of the 5 β -diketol 17". Several experiments were run with regular efficient conversion rates. In some cases, up to 80% of the labeled 5 β -diketol 17" incubated was converted into a mixture of ecdysone and 3-dehydroecdysone in similar proportions.

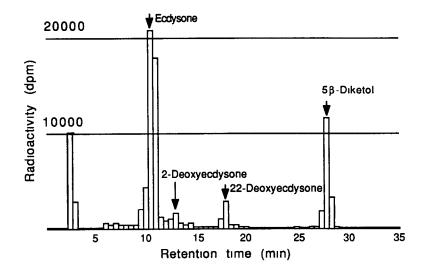


Figure 1 Conversion of trutated 14α -hydroxy-5 β -cholest-7-en-3,6-dione (5 β -diketol) by larval prothoracic glands of *Locusta migratoria* Five pairs of prothoracic glands were excised from 5-day old fifth instar larvae, incubated for 22h with 0.2 μ M trutated 5 β -diketol after which the radioactive molecules were extracted with methanol and separated on a C-18 reverse phase HPLC column, elution with a gradient from 10% to 100% methanol in water over 30 min Columns radioactivity measurements of aliquots of each fraction Arrows position of reference molecules detected by U V absorbance

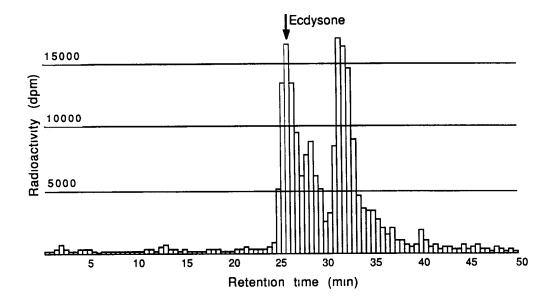


Figure 2 Radiochromatogram of the compound which comigrate with ecdysone, injected together with reference molecule on a C-18 reverse phase HPLC column, elution with a gradient from 15% to 20% acetonitrile in water over 20 min Columns radioactivity measurements of each fraction Arrow position of reference ecdysone detected by U V absorbance

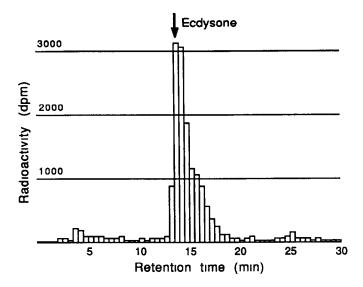


Figure 3 Radiochromatogram of the putative 3-dehydroecdysone after incubation with hemolymph, injected together with reference ecdysone on a C-18 reverse phase HPLC column, isocratic elution with a 20% acetonitrile in water as solvent system Columns radioactivity measurements of each fraction Arrow position of reference ecdysone by U V absorbance.

CONCLUSION

Our initial approach to 14α -hydroxy-5 β -cholest-7-ene-3,6-dione (17) involved the synthesis of 5 as an intermediate However, the published procedure did not give the desired molecule and it was necessary to synthesize 5 by a different method. As a result, our modified synthetic route is novel and also shorter than the original pathway. The tritiated molecule 17" was very efficiently converted to ecdysone (1) and 3-dehydroccdysone (2). This labelled molecule appears to be a potent precursor in ecdysteroid biosynthesis. This first result should now allow investigations into the role of 3-dehydroccompounds in ecdysteroid biosynthesis.

ACKNOWLEDGMENTS

The authors wish to thank Mr Jean Daniel Sauer and Mrs Elisabeth Krempp for NMR spectra and Dr Gérard Teller for mass spectra. We express our gratitude to Pr Guy Ourisson for valuable discussions and for critical reading of this manuscript.

EXPERIMENTAL

Melting points were measured on a Reichert hot stage microscope and are uncorrected $[\alpha]_D$ were measured on a Perkin-Elmer 141 polarimeter in CHCl₃ IR spectra were recorded in KBr on a Perkin-Elmer 881 infrared spectrophotometer UV spectra were measured on a Kontron-Uvikon 810 UV-vis spectrophotometer NMR spectra were recorded on a Bruker SY (200 MHz) apparatus with CHCl₃ ($\delta = 7.26$ ppm) as internal standard for ¹H NMR, CDCl₃ ($\delta = 77.02$ ppm) as internal standard for ¹³C NMR and CHCl₃ ($\delta = 7.26$ ppm) as internal standard for ²H NMR. The chemical shifts are reported in ppm downfield from TMS (*, + = interchangeable assignment) MS were measured on a LKB 9000 S apparatus by direct introduction, or coupled to a GC DB5 column (J W), an ionization potential of 70 eV was used Microanalyses were performed by the Strasbourg Division of the Service Central de Microanalyse of CNRS TLC were run on pre-coated plates of silica gel 60 F 254 (Merck), dipped in a solution of vanillin (1 g) in EtOH/H₂SO₄ (95/5, 1 l) and heated on a hot plate to reveal the compounds Medium pressure chromatography (P = 0.5 - 1.1 bar) was conducted on silica gel (40 - 63 mm, Merck) columns Radioactivity has been determined with a Kontron Betamatic V counter (equipped with external standards) All solvents were freshly distilled before use Air- or moisture-sensitive reactions were conducted in flame-dried glassware and under an inert atmosphere

FIRST PATHWAY

Cholesta-4,7-dien-3-one (7)

A toluene solution (400 ml, freshly distilled) containing 7-dehydrocholesterol (6) (20 0 g, 52 0 mmol), cyclohexanone (60 ml, 0 58 mol, freshly distilled) and 10 0 g of molecular sieve 0 4 nm was stirred at room temperature for 2 hours. After the rapid addition of 6 0 g (29 3 mmol) of aluminium isopropoxide, the orange-red reaction solution was stirred and boiled under reflux for 30 minutes. The solution was cooled to room temperature, washed successively with cold (5°C) 2N hydrochloric acid, cold water, cold

aq sat. sodium hydrogen-carbonate and cold brine, dried with sodium sulfate and filtered The toluene solution was concentrated to dryness under reduced pressure and the residue was chromatographed on silica gel Elution with hexane/ethyl acetate 95/5 gave cholesta-4,7-dien-3-one (7), 17 0 g (85%)

7 Mp 87-89°C (lit.⁷ mp 87-89°C), $[\alpha]_D 23 + 33$ (c = 1 0), UV λ_{max} (ethanol) 238 nm (ϵ = 15500), IR v (cm⁻¹) 1690, 1630, 1470, ¹H NMR 200 MHz (CDCl₃) δ 0 59 (s, 3H, H-18), 0 86⁺ (d, J = 6 61 Hz, 3H, H-26), 0 87⁺ (d, J = 6 59 Hz, 3H, H-27), 0 93 (d, J = 6 26 Hz, 3H, H-21), 1 17 (s, 3H, H-19), 5 18 (m, w_{1/2} = 9 0 Hz, 1H, H-7), 5 79 (d, J = 1 72 Hz, 1H, H-4), SM m/z 382 (100 0) (M⁺, C₂₇H₄₂O), 367 (9 0), 339 (10 8), 338 (37 9), 259 (20 6), 247 (11 6), 227 (10 5), 136 (20 7), Microanalysis calc for C₂₇H₄₂O (382 3578) C 84 81, H 11 07, found C 84 78, H 11 18, ¹³C NMR n table 1

Cholesta-4,6,8(14)-trien-3-one (8)

First synthesis

A dioxane solution (10 mi) containing cholesta-4,7-dien-3-one (7) (100 0 mg, 0 26 mmol) and DDQ (2,3-dichloro-5,6dicyano-benzoquinone, 123 9 mg, 0 55 mmol, 2 1 eq) was refluxed during 4 hours. After filtration of the hydroquinone, the solution was washed with aq 1% sodium hydroxyde solution and brine, dried with sodium sulfate and filtered. The solution was concentrated to dryness under reduced pressure. Cristallisation from acetone gave cholesta-4,6,8(14)-trien-3-one (8), 55 0 mg (55%)

8 Mp 61-62°C (lut.¹² mp 61-63°C), $[\alpha]_D^{20}$ + 635 (c = 1 2), UV λ_{max} (CH₃CN) 240 nm (ϵ = 4500), 282 nm (ϵ = 8000), 348 nm (ϵ = 25500), IR v (cm⁻¹) 2946, 2872, 1662, 1584, 1266, 1220, 1196, ¹H NMR 200 MHz (CDCl₃) δ 0 87 (d, J = 6 62 Hz, 6H, H-26,27), 0 96 (d, J = 6.51 Hz, 3H, H-21), 0 95 (s, 3H, H-19), 0 99 (s, 3H, H-18), 5 73 (s, 1H, H-4), 6 03* (d, J = 9 29 Hz, 1H, H-7), 6 62* (d, J = 9 29 Hz, 1H, H-6), SM m/z 380 (68 6) (M⁺, C₂₇H₄₀O), 365 (17 6), 268 (100 0), 214 (25 9), Microanalysis calc for C₂₇H₄₀O (380 6120) C 85 20, H 10 59, found C 85 22, H 10 51, 1³C NMR in table 1

Second synthesis

To a tetrahydrofuran solution (10 ml) containing cholesta-4,7-dien-3-one (7) (100 0 mg, 0 26 mmol) was added dropwise at 0°C tri-N-methylanilinium perbromide (108 0 mg, 0 28 mmol, 1 1 eq) in tetrahydrofuran (8 ml) After addition (5 minutes), the mixture was sturred 15 minutes at 0°C and at room temperature for a further 15 minutes. The mixture was then poured into water and the product isolated with ether. The organic layer was washed with water, brine dried with sodium sulfate and evaporated to dryness. The residue was chromatographed on silica gel. Elution with hexane/ethyl acetate 95/5 gave cholesta-4,6,8(14)-trien-3-one (8), 63 1 mg (63%).

SECOND PATHWAY

5α-Cholest-7-en-3β-ol (9)

7-dehydrocholesterol (6) (100 g, 260 mmol) was dissolved in a 1/1 (v/v) mixture of toluene and ethyl acetate Hydrogenation was conducted over palladium (5%) on activated charcoal (15 g) at room temperature and atmospheric pressure during 2 hours The solution was filtered on celite and evaporated to dryness to give 5 α -cholest-7-en-3 β -ol (9), 9.85 g (98%)

9 ¹H NMR 200 MHz (CDCl₃) δ 0 53 (s, 3H, H-18), 0 79 (s, 3H, H-19), 0 86 (d, J = 6 56 Hz, 6H, H-26,27), 0 94 (d, J = 5 97 Hz, 3H, H-21), 3 64 (m, w_{1/2} = 25 0 Hz, 1H, H-3), 5 16 (m, w_{1/2} = 6 7 Hz, 1H, H-7), SM m/z 386 (100 0) (M⁺, C₂₇H₄₆O), 371 (23.6), 273 (14 4), 255 (47 1), 231 (14 9), 228 (14 4), 213 (15 5), Microanalysis calc for C₂₇H₄₆O (386 6594) C 83 87, H 11 99, found C 83 82, H 11 75, ¹³C NMR in table 1

5a-Cholest-7-en-3-one (10)

To a solution of 5α -cholest-7-en-3 β -ol (9) (7 5 g, 19 4 mmol) in dry methylene chloride (200 ml) was added 9 0 g of pyridinium-chlorochromate absorbed on alumina. After stirring for 12 hours, the solid is filtered on alumina and washed with ethyl acetate. The combined filtrates were evaporated to dryness to afford pure 5 α -cholest-7-en-3-one (10), 7 1 g (95%)

10 Mp 144-146°C (ltt.¹² mp 144-146°C), $[\alpha]_D^{20} + 25$ (c = 3 9), IR v (cm⁻¹) 1725, 1665, ¹H NMR 200 MHz (CDCl₃) δ 0 56 (s, 3H, H-18), 0 87 (d, J = 6 53 Hz, 6H, H-26,27), 0 93 (d, J = 6 31 Hz, 3H, H-21), 1 01 (s, 3H, H-19), 5 19 (b, w_{1/2} = 9 0 Hz, 1H, H-7), SM m/z 384 (100 0) (M⁺, C₂₇H₄₄O), 369 (31 1), 272 (15 3), 271 (65 5), 245 (15.2), 244 (16 5), 229 (44 3), Microanalysis calc for C₂₇H₄₄O (384 6436) C 84 31, H 11 53, found C 83 12, H 11 90, ¹³C NMR in table 1

2a,4a-Dibromo-5a-cholest-7-en-3-one (11)

A solution of 5α -cholest-7-en-3-one (10) (1 0 g, 2 6 mmol) in tetrahydrofuran (20 ml) containing tri-N-methylanilinium perbromide (2 0 g, 5 3 mmol, 2 1 eq) was stured during 15 minutes at 0°C and at room temperature for a further 15 minutes. The mixture was then poured into water and the product isolated with ether. The organic layer was washed with water, brine dried with sodium sulfate and evaporated to dryness to yield 2α , 4α -dibromo- 5α -cholest-7-en-3-one (11), 630 mg (64%), which was normally used directly for the next reaction (dielimination). Recristallization from methanol/ether of an aliquot gave 2α , 4α -dibromo- 5α cholest-7-en-3-one as white needles

11 Mp 195-197°C (ht.¹² mp 192-195°C), $[\alpha]_{D}^{20}$ - 20 (c = 0 9), UV λ_{max} (CH₃CN) 197 nm (ε = 13200), IR v (cm⁻¹) 2962, 2844, 1741, 1455, 1388, 1163, ¹H NMR 200 MHz (CDCl₃) δ 0 57 (s, 3H, H-18), 0 87 (d, J = 6 49 Hz, 6H, H-26,27), 0 91 (d, J = 6 14 Hz, 3H, H-21), 1 20 (s, 3H, H-19), 2 55 (b, w_{1/2} = 26 3 Hz, 1H, H-6_{eq}), 2 75 (dd, J = 13 16 et 5 26 Hz, 1H, H-1_{eq}), 4 67 (d, J = 13 16 Hz, 1H, H-4_{ax}), 4 87 (dd, J = 14 21 et 5 78 Hz, 1H, H-2_{ax}), 5 25 (bd, w_{1/2} = 11 6 Hz, 1H, H-7). SM m/z 544 (53 1), 543 (31 0), 542 (100 0) (M⁺, C₂₇H₄₂OBr₂), 462 (39 5), 431 (47 1), 429 (93 1), 427 (49 4), 387 (41 1), 381 (40 6), 323 (57 5), Microanalysis calc for C₂₇H₄₂OBr₂ (542 4358) C 59 78, H 7 80, found C 59 60, H 7 60, ¹³C NMR in table 1

Cholesta-1,4,7-trien-3-one (4)

A solution of the unpurified dibromide 11 (3 0 g, 5 5 mmol) in dimethylformamide (20 ml) containing lithium carbonate (5 0 g) and lithium bromide (2 5 g) was refluxed during 30 minutes. The solution was cooled to room temperature, washed with water and brine, dried with sodium sulfate and evaporated to dryness. The residue was chromatographed on silica gel. Elution with hexane/ethyl acetate 95/5 gave cholesta-1,4,7-trien-3-one (4), 2 7 g (90%).

4 Mp $130-132 \circ C$ (ltt.¹² mp $130-132^{\circ}C$), $[\alpha]_D^{20} - 15$ (c = 28), UV λ_{max} (CH₃CN) 243 nm (ε = 14800), IR v (cm⁻¹) 1650, 1625, 1600, ¹H NMR 200 MHz (CDCl₃) δ 0 63 (s, 3H, H-18), 0 87 (d, J = 6 48 Hz, 6H, H-26,27), 0 94 (d, J = 6 28 Hz, 3H, H-21), 1 24 (s, 3H, H-19), 2 81 (d, J = 18 80 Hz, 1H, H-6), 3 17 (d, J = 18 80 Hz, 1H, H-6), 5.25 (m, w_{1/2} = 10 0 Hz, 1H, H-7), 6 13 (m, w_{1/2} = 5 0 Hz, 1H, H-4), 6.26 (dd, J = 9 99 Hz, 1H, H-2), 7 09 (d, J = 10 05 Hz, 1H, H-1), SM m/z 380 (100 0) (M⁺, C₂₇H₄₀O), 365 (28 0), 268 (31 2), 172 (48 9), 157 (59 3), 133 (64 8), 121 (59 0), 105 (42 0), Microanalysis calc for C₂₇H₄₀O (380 6120) C 85 20, H 10 59, found C 84 93, H 10 78, ¹³C NMR in table 1

Cholesta-4,7-dien-3-one (7)

Cholesta-1,4,7-trien-3-one (4) (200 0 mg, 0 53 mmol) was dissolved in ethanol (10 ml) After addition of 20 ml of quinoline, the hydrogenation was conducted over palladium (5%) on calcium carbonate (20 mg) at room temperature during 16 minutes (1 Atm pressure) The solution was immediately filtered and evaporated to dryness. The residue was chromatographed on

silica gel Elution with hexane/ethyl acetate 97/3 gave a mixture of 5 β -cholest-7-en-3-one (12), 350 mg (17%), cholesta-4,7-dien-3-one (7), 1200 mg (60%) and starting material 4, 200 mg (10%)

The previous procedure, without quinoline or using Palladium (5%) on activated charcoal, gave the same products, but with different ratio

5β -Cholest-7-en-3-one (12)

Cholesta-1,4,7-truen-3-one (4) (200 0 mg, 0.53 mmol) was dissolved in ethanol (10 ml) After addition of 20 ml of quinoline, the hydrogenation was conducted over palladium (5%) on calcium carbonate (20 mg) at room temperature during 30 minutes. The solution was immediately filtered and evaporated to dryness. The residue was chromatographed on silica gel Elution with hexane/ethyl acetate 97/3 gave 5 β -cholest-7-en-3-one (12), 179 0 mg (90%).

The previous procedure, without quinoline or using Palladium (5%) on activated charcoal, gave the same product

12 Mp 87-88°C (lut.²³ mp 87-88°C), $[\alpha]_D^{23}$ + 65 (c = 0 57), IR v (cm⁻¹) 1725, 1665, ¹H NMR 200 MHz (CDCl₃) δ 0 57 (s, 3H, H-18), 0 87 (d, J = 6 61 Hz, 6H, H-26,27), 0 93 (d, J = 6 24 Hz, 3H, H-21), 0 99 (s, 3H, H-19), 5 12 (m, w_{1/2} = 10 0 Hz, 1H, H-7), SM m/z 384 (92 4) (M⁺, C₂₇H₄₄O), 369 (20 4), 352 (30 0), 351 (100 0), 334 (30 3), 333 (49 8), 119 (32 6), 105 (68 0). Microanalysis calc for C₂₇H₄₄O (384 6436) C 84 31, H 11 53, found C 83 99, H 11 62, ¹³C NMR in table 1

THIRD PATHWAY

5β-Cholest-7-en-3-one (12)

A solution of cholesta-4,7-dien-3-one (7) (2 0 g, 5 2 mmol), 60 ml of ethanol, 12 ml of aq 0 1N sodium hydroxide and 200 0 mg of palladium (5%) on activated charcoal (150 mg) was shaken with hydrogen at room temperature and atmospheric pressure for 2 hours. The catalyst was removed by filtration The solution was washed with water, brine and then dried with sodium sufate and evaporated to dryness. The residue was chromatographed on silica gel Elution with hexane/ethyl acetate 95/5 gave 1 4 g of 5 β -cholest-7-en-3-one (12), (72%) and 150 0 mg of 5 α -cholest-7-en-3-one (10), (7%)

5β -cholesta-1,7-dien-3-one (5)

First step

A solution of 5 β -cholest-7-en-3-one (12) (435 0 mg, 1 1mmol) in tetrahydrofuran (20 ml) containing tri-N-methylanilinium perbromide (468 0 mg, 1 24 mmol, 1 1 eq) was sturred during 15 minutes at 0°C and at room temperature for a further 15 minutes The mixture was then poured into water and the product isolated with ether. The organic layer was washed with water, brine, dried with sodium sulfate and evaporated to dryness to yield 2 α -bromo-5 β -cholest-7-en-3-one (13), 350 mg (80%) (which was normally used directly for the next reaction). Cristallisation from methanol/ether of an aliquot gave 2 α -bromo-3,3-dimethoxy-5 β -cholest-7-ene (14) in white needles

14 Mp $124-126^{\circ}C$, $[\alpha]_{D}^{23}$ + 22 (c = 0 89), IR v (cm⁻¹) 2960, 2918, 2875, 1458, 1378, 1119, 1098, 1053, ¹H NMR 200 MHz (CDCl₃) δ 0 53 (s, 3H, H-18), 0 87 (d, J = 6 62 Hz, 6H, H-26,27), 0 90 (s, 3H, H-19), 0 92 (d, J = 6 14 Hz, 3H, H-21), 2 22 (dd, J = 14 50 et 5 00 Hz, 1H, H-1_{eq}), 2 41 (b, w_{1/2} = 20 00 Hz, 1H, H-6_{eq}), 3 34 (s, 3H, H-1'), 3 40 (s, 3H, H-1''), 4 34 (dd, J = 12 97 et 4 46 Hz, 1H, H-2_{ax}), 5 10 (bd, w_{1/2} = 10 0 Hz, 1H, H-7), SM m/z 510 (4 1), 508 (3 0) (M⁺, C₂₉H₄₉O₂Br), 478 (22 3), 476 (21 9), 398 (32 6), 397 (100 0), 366 (28 3), 365 (85 1), 314 (21 2), 313 (57 9), 279 (14 1), 256 (30 6), 242 (15 9), 228 (19 5), Microanalysis calc for C₂₉H₄₉O₂Br (509 6081) C 68 35, H 9 69, found C 68 19, H 9 40, ¹³C NMR in table 1

F DOLLE et al

Second step

A solution of the unpurfied bromide 13 (350 0 mg, 0 76 mmoi) in dimethylformamide (15 ml) containing lithium carbonate (14 g) and lithium bromide (750 0 mg) was refluxed during 30 minutes. The solution was cooled to room temperature, washed with water and brine, dried with sodium sulfate and evaporated to dryness. The residue was chromatographed on silica gel Elution with hexane/ethyl acetate 95/5 gave a mixture of 5 β -cholesta-1,7-dien-3-one (5), 283 1 mg (80 1%), cholesta-4,7-dien-3-one (7), 27 5 mg (7 8%) and cholesta-1,4,7-trien-3-one (4), 11 7 mg (3 3%)

5 Mp 93-95 °C, $[\alpha]_D^{23}$ + 41 (c = 10), UV λ_{max} (CH₃CN) 196 nm (ϵ = 14100), 230 nm (ϵ = 11100), IR v (cm⁻¹) 2959, 2934, 2875, 1667; 1466, 1442, 1412, 1372, ¹H NMR 200 MHz (CDCl₃) δ 0 57 (s, 3H, H-18), 0 86 (d, J = 6 54 Hz, 6H, H-26,27), 0 92 (d, J = 6 29 Hz, 3H, H-21), 1 26 (s, 3H, H-19), 5.15 (b, w_{1/2} = 10 0 Hz, 1H, H-7), 5 88 (d, J = 10 00 Hz, 1H, H-2), 6 98 (d, J = 10.03 Hz, 1H, H-1), SM m/z 382 (87 4) (M⁺, C₂₇H₄₂O), 275 (23 3), 274 (100.0), 269 (16 1), 161 (22.5), 109 (25 4), Microanalysis calc for C₂₇H₄₂O (382 6278) C 84 75, H 11 06, found C 84 92, H 11 18, ¹³C NMR in table 1

5B-Cholesta-1,7-diene-3,6-dione (15)

In 10 ml of methylene chloride at room temperature and under a nitrogen atmosphere, 100 0 mg (0 26 mmol) of 5 β -cholesta-1,7-dien-3-one (5) was dissolved To the sturred solution was added 700 mg (2 6 mmol) of CrO₃-(pyridine)₂ complex (Collins reagent, freshly prepared) The mixture immediately began turning brown and depositing a tarry precipitate on the sides and bottom of the flask. After 15 hours of stirring at room temperature, an additional 700 mg (2.6 mmol) of CrO₃-(pyridine)₂ complex was added to the reaction mixture 4 hours later, thin layer analysis of the solution showed the presence of a trace of starting material (longer oxidation periods did not eliminate this product) The reaction mixture was poured from the flask. The precipitate remaining in the flask was rinsed with small portions of ether (little improvement in yield was found when the tarry precipitate was dissolved in aq. sat. sodium hydrogen-carbonate and then extracted with ether) The organic layer was washed several times with aq sat. sodium hydrogen-carbonate, 5% hydrochloric acid, dried with sodium sulfate and filtered. The solvents were evaporated to dryness and the residue was chromatographed on silica gel. Elution with hexane/ethyl acetate 95/5 gave 5 β -cholesta-1,7-diene-3,6-dione (15), 27 0 mg (27%)

15 Mp 122-124°C, $[\alpha]_D^{23}$ + 33 (c = 1 02), UV λ_{max} (CH₃CN) · 194 nm (ε = 13900), 225 nm (ε = 18700), IR v (cm⁻¹) 2927, 2867, 1654, 1455, 1380, 1262, ¹H NMR 200 MHz (CDCl₃) δ 0 62 (s, 3H, H-18), 0.87 (d, J = 6 55 Hz, 6H, H-26,27), 0 93 (d, J = 5.88 Hz, 3H, H-21), 1 26 (s, 3H, H-19), 5 77 (bs, w_{1/2} = 5 0 Hz, 1H, H-7), 5 95 (d, J = 10.17 Hz, 1H, H-2), 6 79 (d, J = 10 18 Hz, 1H, H-1), SM m/z 396 (85.5) (M⁺, C₂₇H₄₂O₂); 381 (19 0), 378 (23 1), 288 (22 1), 284 (23 0), 183 (100.0), 185 (24.5), Microanalysis calc for C₂₇H₄₀O₂ (396 6110) C 81 77, H 10 16, found C 81 95, H 10 03, ¹³C NMR in table 1.

14α-Hydroxy-5β-cholesta-1,7-diene-3,6-dione (16)

To a solution of 250 0 mg (0 63 mmol) of 5 β -cholesta-1,7-diene-3,6-dione (15) in freshiy distilled dioxan (20 ml), was added 700 mg (6.31 mmol, 10 eq) of finely powdered SeO₂ and the mixture was refluxed during 15 minutes. The solution was then cooled to 0°C and allowed to warm to room temperature. After filtration of the solution on celute, the solvent was removed. The residue was chromatographed on silica gel. Elution with hexane/ethyl acetate 92/8 gave 14 α -hydroxy-5 β -cholesta-1,7-diene-3,6-dione (16), 197 6 mg (76%) and 14 α -hydroxy-cholesta-1,4,7-tinene-3,6-dione (18), 25 0 mg (10%)

16 Mp 179-181°C, $[\alpha]_{D}^{23}$ + 73 (c = 0.75), UV λ_{max} (CH₃CN) : 194 nm (ε = 10700), 224 nm (ε = 19400), IR v (cm⁻¹) 3457, 2954, 2908 ; 1662 ; 1456, 1379, ¹H NMR 200 MHz (CDCl₃) δ . 0 71 (s, 3H, H-18), 0 87 (d, J = 6 52 Hz, 6H, H-26,27), 0.93 (d, J = 6 22 Hz, 3H, H-21), 1.27 (s, 3H, H-19), 3 14 (bt, w_{1/2} = 20.0 Hz, 1H, H-9), 5 92 (d, J = 2 10 Hz, 10 Hz)

1H, H-7), 5 95 (d, J = 10 03 Hz, 1H, H-2), 6 82 (d, J = 10 05 Hz, 1H, H-1), SM m/z 412 (34 1) (M⁺, C₂₇H₄₀O₃), 394 (51 0), 277 (27 3), 276 (100 0), 229 (42 9), 109 (34 4), Microanalysis calc for $C_{27}H_{40}O_3$ (412 6100) C 78 59, H 977, found C 78 78, H 9.50, ¹³C NMR in table 1

18 Mp 95-97°C, ¹H NMR 200 MHz (CDCl₃) δ 0 78 (s, 3H, H-18), 0 88 (d, J = 6 51 Hz, 6H, H-26,27), 0 94 (d, J = 6 17 Hz, 3H, H-21), 1 34 (s, 3H, H-19), 3 06 (bt, w_{1/2} = 20 0 Hz, 1H, H-9), 6 17 (bs, w_{1/2} = 5 0 Hz, 1H, H-7), 6 33 (d, J = 10 20 Hz, 1H, H-1), 6 68 (bs, w_{1/2} = 5 0 Hz, 1H, H-4), 7 06 (d, J = 9 82 Hz, 1H, H-2), SM m/z 410 (66 8) (M⁺, C₂₇H₃₈O₃), 382 (38 4), 228 (82 7), 227 (72 4), 200 (34 1), 135 (97 0), Microanalysis calc for C₂₇H₃₈O₃ (410 5942) C 78 98, H 9 33, found C 79 03, H 9 40, ¹³C NMR in table 1

14α -Hydroxy-5 β -cholest-7-ene-3,6-dione (17)

The compound 16 (100 0 mg, 0 24 mmoi) was dissolved in ethyl acetate. Hydrogenation was conducted over palladium (5%) on activated charcoal (20 0 mg) at room temperature and atmospheric pressure during 1 hour. The solution was filtered and evaporated to dryness to give 14α -hydroxy-5 β -cholest-7-ene-3,6-dione (17), 98 mg (98%)

17 Mp 172-174°C, $[α]_D^{23}$ + 75 (c = 0 92), UV λ_{max} (CH₃CN) 194 nm (ε = 11800), 243 nm (ε = 14700), IR v (cm⁻¹) 3502, 2964, 1720, 1643, 1462, 1378, 1318, 1253, ¹H NMR 200 MHz (CDCl₃) δ 071 (s, 3H, H-18), 0 87 (d, J = 6 52 Hz, 6H, H-26,27), 0 93 (d, J = 6 22 Hz, 3H, H-21), 1 09 (s, 3H, H-19), 3 27 (bt, w_{1/2} = 20 0 Hz, 1H, H-9), 5 87 (d, J = 2 18 Hz, 1H, H-7), SM m/z 414 (87 3) (M⁺, C₂₇H₄₂O₃), 386 (67 6), 234 (76 2), 232 (79 7), 231 (100 0), Microanalysis calc for C₂₇H₄₂O₃ (414 6258) C 78 21, H 10 21, found C 77 90; H 10 14, ¹³C NMR in table 1

$[1,2-^{2}H_{2}]$ -14 α -Hydroxy-5 β -cholest-7-ene-3,6-dione (17')

The procedure described above, under deuterium atmosphere (1 Atm pressure, with 7 5 mg of 14 α -Hydroxy-5 β -cholesta-1,7-diene-3,6-dione (16)), gave [1,2-²H₂]-14 α -hydroxy-5 β -cholest-7-ene-3,6-dione (17') quantitatively (100%)

17' ¹H NMR 200 MHz (CDCl₃) δ = 0 71 (s, 3H, H-18), 0 87 (d, J = 6.52 Hz, 6H, H-26,27), 0 93 (d, J = 6 22 Hz, 3H, H-21), 1 09 (s, 3H, H-19), 3 27 (bt, $w_{1/2} = 200$ Hz, 1H, H-9), 5 87 (d, J = 2 18 Hz, 1H, H-7), ²H NMR 61 4 MHz (CHCl₃) δ 1 52 (b, $w_{1/2} = 101$ Hz, 1D, D-1), 2 33 (b, $w_{1/2} = 123$ Hz, 1D, D-2), SM m/z 417 (17 9), 416 (56 7) (M⁺, C₂₇H₄₀O₃D₂), 415 (45 5), 389 (17 0), 388 (51 6), 236 (49 1), 235 (61 8), 234 (63 5), 233 (100 0), 232 (62 8), 174 (20 8), 161 (29 1)

$[1,2-^{2}H_{2}]-3\alpha,14\alpha$ -Dihydroxy-5 β -cholest-7-en-6-one (19)

To a solution of $[1,2-^{2}H_{2}]-14\alpha$ -hydroxy-5 β -cholest-7-ene-3,6-dione (17') (2 0 mg, 4 8 mmol) in methanol (1 ml) was added 2 0 mg of sodium borohydride. The mixture was sturred 5 minutes, then 1 ml of water was added. Extraction with ether gave pure $[1,2-^{2}H_{2}]-3\alpha,14\alpha$ -dihydroxy-5 β -cholest-7-ene-6-one (19), 1 5 mg (75 %)

19 ¹H NMR 200 MHz (CDCl₃) δ 0 68 (s, 3H, H-18), 0 87 (d, J = 6 51 Hz, 6H, H-26,27), 0 93 (d, J = 6 45 Hz, 3H, H-21), 1 23 (s, 3H, H-19), 3 18 (bt, w_{1/2} = 20 0 Hz, 1H, H-9), 3 66 (td, J = 10 78 et 3 53 Hz, 1H, H-3), 5 87 (d, J = 2 45 Hz, 1H, H-7), SM m/z 418 (21 9) (M⁺, C₂₇H₄₂O₃D₂), 400 (41 2), 399 (25 0), 390 (45 8), 389 (26.8), 372 (28 7), 235 (26 4), 217 (21 7)

[1,2-³H₂]-14α-Hydroxy-5β-cholest-7-ene-3,6-dione (17")

The procedure described above was carried out under a tritium atmosphere at the CEA (Saclay)

Compound 16 (2.5 mg, 8.0 µmol) was dissolved in ethyl acetate (2 ml) Tritiation (tritium gas 0.67 TBq, 20 Ci) was conducted (via a Toepler pump) over palladium (5%) on activated charcoal (4.0 mg) at room temperature and at 500 mmHg pressure

during 40 mn The solution was filtered and evaporated to dryness. The labile tritium was eliminated by dilution in methanol and evaporation, twice. The compound 17" (6.9 GBq, 186 mCi) was obtained with a radiochemical purity about 96% by High Performance Liquid Chromatography (HPLC, silica column, hexane/ethyl acetate 77/23). The analytical radio-chromatography was made on an reverse phase column in analytical HPLC (C-18 ODS, methanol/water 90/10).

17" ³H NMR 320 MHz (Bruker C300, ¹H decoupled) (DMSO) δ 1 43 (d, J = 1 63 Hz, 1T, T-1), 2 14 (d, J = 1 63 Hz, 1T, T-2), SM (Finnigan 4600, DCI/NH₃) m/z 436 (29 6) ([M+NH₃]⁺, $C_{27}H_{40}O_3T_2 + NH_3$), 434 (13 5), 432 (1 9), Specific activity 1 74 TBq/mmol (47 Ci/mmol)

REFERENCES

- 1 Warren, JT, Sakurai, S, Rountree, DB, Gilbert, LI, Lee, SS, Nakanishi, K Proc Natl Acad Sci USA 1988, 85, 958-962
- 2 Kirushi, S., Rountree, D.B., Sakurai, S., Gilbert, L.I. Experientia 1990, 46, 716-721
- 3 Sakurai, S, Warren, JT, Gilbert, LI Arch Insect Biochem Physiol 1981, 10, 179-197
- 4 Warren, J T, Hetru, C Inv Reprod Dev 1990, 18, 91-100
- 5 Rees, H H In "Ecdysone", Koolman, J ed, Georg Thieme Verlag, Stuttgart 1989, 152-160
- 6 Rubin, M, Armbrecht, H J Am Chem Soc 1953, 75, 3513-3516
- 7 Cohen, C.F., Louloudes, S.J., Thompson, M.J. Steroids 1957, 591-600
- 8 Turner, A B J Chem Soc 1968, 2568-2570
- 9 Walker, D., Hiebert, J Chem Rev 1967, 67, 153-195
- 10 Brynjolffssen, J., Hands, D., Midgley, J.M., Whalley, W.B. J. Chem. Soc. 1975, 826-828
- 11 Cheng, Y S , Liu, W L , Chen, S H Synthesis 1980, 223-224
- 12 Emke, A, Hands, D, Midgley, JM, Whalley, WB, Ahmad, R J Chem Soc Perkan Trans I 1976, 820-822
- 13 Kunzer, H, Stahnke, M, Sauer, G, Wiechert, R Tetrahedron Letters 1990, 27, 3859-3862
- 14 Velasco, M, Rivera, J, Rosenkanz, G; Sendheimer, F, Djerassi, C J Org Chem 1953, 18, 92-95
- 15 Shimahara, M, Nishimura, S Chemistry and industry 1966, 1796-1797
- 16 Nishimura, S., Shimahara, M., Shiota, M. J. Org. Chem. 1966, 31, 2394-2395
- 17 Dauben, WG, Lorber, M, Fullerton, DS J Org Chem 1969, 34, 3587-3592
- 18 Hetru, C, Nakatani, Y, Luu, B Nouv J Chum 1983, 7, 587-591
- 19 Joyce, M.J., Hiremath, S.V., Mattammal, M.B., Elliott, W.H. Steroids 1984, 44, 95-101
- 20 Tal, D M, Elliott, W H J Labelled Compd Radiopharm 1985, 22, 359-366
- 21 Haag, T, Meister, MF, Hetru, C, Kappler, C, Nakatani, Y, Beaucourt, JP, Rousseau, B, Luu, B Insect Biochem 1987, 17, 291-301
- 22 Lafont, R, Sommé-Martin, G, Mauchamp, B, Maume, BF, Delbecque, JP In "Progress in Ecdysone Research", Hoffmann, JA ed, Elsevier, Amsterdam 1980, 45-68
- 23 Morand, PG, Henbest, HB, Jackson, WR J Chem Soc C, 1967, 2467-2473